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Bob-I is expressed in classic Hodgkin lymphoma

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Abstract

Background: Almost all researchers agree on the lack of Bob-I expression in Hodgkin/Reed-Sternberg (H/RS) cells in classic Hodgkin lymphoma (CHL), and utilize this marker as a diagnostic tool in conjunction with other markers to differentiate between lymphocyte predominance Hodgkin lymphoma (LPHL) and CHL.

Aim: To study the immunohistochemical (IHC) expression of Bob-I in Egyptian CHL and to correlate this expression with Epstein-Barr virus (EBV) viral load.

Materials and methods: Paraffin sections of randomly selected 18 CHL cases were included: 2 lymphocyte rich (LR), 4 mixed cellularity (MC), 10 nodular sclerosis (NS) and 2 lymphocyte depletion (LD). All cases were immunostained for Bob-I. EBV was evaluated by EBV early RNA transcripts in situ hybridization (EBER ISH) and immunostaining for EBV latent membrane protein-1 (LMP-1).

Results: Sixty seven percent of cases (12/18) were positive for EBV by ISH and/or immunostaining for LMP-1. Moderate to strong nuclear Bob-I was observed in 94% of cases. The positivity ranged between 25–100%. Bob-I immunoreactivity was strongly associated with EBV positivity ($p < 0.001$).

Conclusion: This study proves nuclear IHC expression of Bob-I on H/RS in CHL implying the difficulties in applying this marker to differentiate between LPHL and CHL. Does this difference between Western and Egyptian CHL reflect genetic and/or environmental factors, or simply no difference exists as most researchers are concentrated on the Western population and no comparative studies have been done. Studies from other countries might answer this question.

Background

According to the World Health Organization (WHO) classification of haematological malignancies [1], the B-cell specific transcriptional co-activator or Bob-1/OCA-B is not expressed on H/RS cells, a point used to differentiate between CHL and LPHL, the latter being Bob-1 positive. Bob-1/OCA-B located on chromosome 11q23.1 [2], is involved in the transcription of immunoglobulin (Ig) genes through recruitment of the highly conserved

octamer site of Ig promoters, mediated by either Oct-1 or Oct-2 transcription factor [3]. Bob-1 is essential for the response of B cells to antigens, and is required for the formation of the germinal centre (GC) [4].

EBV, the main etiologic agent for HL [5], has been reported in 30–50% of cases in developed countries and in up to 95% of cases in developing ones [6]. The detection of EBV in H/RS cells is mainly based on the detection

of the latently expressed gene LMP-1 or on the detection of EBV early RNA transcripts (EBER) [7].

As H/RS cells originate from the GC [8-10], they are expected to express B cell markers such as Bob-1. Therefore this work was undertaken to study the expression of Bob-1 in Egyptian CHL and to correlate this expression with EBV viral load in an attempt to find out if differences do exist between Western and Egyptian CHL, for optimal assessment of the treatment regime.

Materials and methods

Classic Hodgkin lymphoma cases

Paraffin sections of randomly selected CHL were classified according to the WHO [1]. A total of 18 CHL cases were studied. These included 2 cases of LRHL, 4 MCHL, 10 NSHL, 2 LDHL. The positive controls consisted of 2 reactive nodes selected with follicular hyperplasia and 2 cases of LPHL. The initial diagnostic panel included CD30, CD15, CD20 and CD3. EMA and ALK-1 were performed whenever indicated to rule out LPHL and anaplastic large cell lymphoma.

Immunohistochemistry

The expression of Bob-1 and EBV LMP-1 was evaluated by immunostaining (table 1). Following deparaffinization, endogenous peroxidase was inhibited by tissue sections incubation for 10 minutes at room temperature in 0.3% H₂O₂. Following antigen retrieval, slides were rinsed in distilled water and finally phosphate buffered saline (PBS). All incubations were performed at room temperature. After incubation with the primary antibody, sections were rinsed in PBS and incubated with the LSAB-2 detection kit and the steps were followed according to the manufacturer instructions (Dakocytomation). Diaminobenzidine tetrachloride (DAB) was applied for 10 minutes and lastly, sections were counterstained with Harris haematoxylin (Hx).

Evaluation of immunohistochemical staining

The percentage of positively stained H/RS cells were semi-quantitatively determined as follows: 0- absence or staining of less than 5% of H/RS cells; +) 5-25%; ++> 25-50%; +++> 50-75%; ++++> 75% of H/RS cells showed

a positive staining. In all cases, small reactive lymphocytes served as positive internal control for Bob-1, in addition to the positivity of the reactive nodes and lymphocytic/histiocytic cells of LPHL.

In-situ hybridization

The ISH steps were performed according to the manufacturer's instructions (Novocastra, NCL-EBV-K). The slides were first dewaxed in xylene, hydrated in descending grades of alcohol and lastly immersed in water. One hundred µl of proteinase K in 0.05 mM Tris/HCL buffer pH 7.6 were applied for 10 minutes at 37°C. This step was followed by slides immersion in water, dehydration and air drying. Depending on the tissue section, 20 to 50 µl of the probe hybridization solution were applied. Sections were coverslipped and incubated for 2 hours at 37°C. The covers were allowed to drain off into a beaker; they were then washed in Tris containing 0.1% triton X-100. For the detection, 100 µl of the blocking solution was applied for 10 minutes followed by rabbit F (ab') anti-FITC conjugated to alkaline phosphatase (AP) diluted 1:100 for 30 minutes. Slides were subsequently washed in TBS followed by the AP substrate buffer. The AP activity was demonstrated by incubation in dark overnight with a mixture solution of 5 bromo-4 chloro indolyl phosphate, nitro-blue tetrazolium (BCNT). Finally, the slides were washed and counterstained with Mayer's haematoxylin. The control of the procedure included positive control sections and negative control probe supplied with the ISH kit.

Evaluation of in-situ hybridisation

The staining was considered positive when dark blue to black dots were encountered in the nuclei of the H/RS cells together with positivity of the positive control tissue and negativity of the duplicate sections hybridized with the negative probe.

Statistical analysis

The association between Bob-1 positive cells and EBV viral load was evaluated using the Chi square test.

Results

In reactive nodes, residual follicles and inflammatory milieu of HL, strong nuclear Bob-1 was expressed in GC B

Table 1: Reagents used for IHC in the study

| | Bob-1 | LMP-1 |
|--------------------------------|--------------------------|--|
| Source | Santa Cruz | Dakocytomation |
| Reference | Sc 955 rabbit polyclonal | M0897, monoclonal |
| Heat induced epitope retrieval | | Citrate pH 6.0, 750 w × 5 m × 3 times |
| Dilution | 1:1500 | 1:50 |
| Incubation | 60 m | 30 m |
| Interpretation | Nuclear | Membranous, cytoplasmic and/or paranuclear dot |

cells while moderate staining was seen in scattered mantle zone B cells and interfollicular T cells.

In CHL, moderate to strong nuclear IHC Bob-1+ H/RS cells were encountered in 94% of cases irrespective of the subtype. The percentage of positive cells ranged from 25–100% (table 2, figures 1, 2, 3). Membranous, cytoplasmic and/or paranuclear dot staining for LMP-1 (figure 4) was identified in 61%, while nuclear staining for EBER-ISH (figure 5) was encountered in 67% of cases. A close association was found between Bob-1 immunoreactivity and EBV viral load ($p < 0.001$).

Discussion

CHD is a B-cell neoplasm in nearly all instances derived from the GC B cells harbouring somatically mutated IgV region genes. However, these cells have consistently lost their Ig gene transcription ability, due to functional defects in the Ig gene regulatory elements [10-12]. The defect has been attributed to crippling mutations [11,12], a defect in the transcription machinery due to lack of expression of the octamer transcription factor Oct2 and/or its coactivator Bob-1 [12,14], or epigenetic silencing in the inhibition of IgH transcription [15].

The expression of Bob-1 in H/RS cells was a surprising finding. To our knowledge, this is the first study reporting strong nuclear Bob-1 in almost all H/RS cells. In reactive nodes and non neoplastic cells in HL, Bob-1 IHC expression is concordant with the literature [16,17]. However, in neoplastic conditions, Bob-1 positivity is restricted to LPHL and has been a useful tool in differentiating it from CHL [13,18-20]. Only one previous study using tissue microarray, reported Bob-1+ H/RS cells in 17% of cases with strong Bob-1 positivity in only 2% of cases [21].

Should the strong IHC expression of Bob-1 in H/RS cells is really to be an unexpected finding? First, in HL cell lines, Oct-2 has been reported in one study to be constantly expressed on H/RS cells [22]. Second, a close resemblance has been found between primary mediastinal B-cell lymphoma (PMBL) and CHL [23-25]. PMBL signature genes

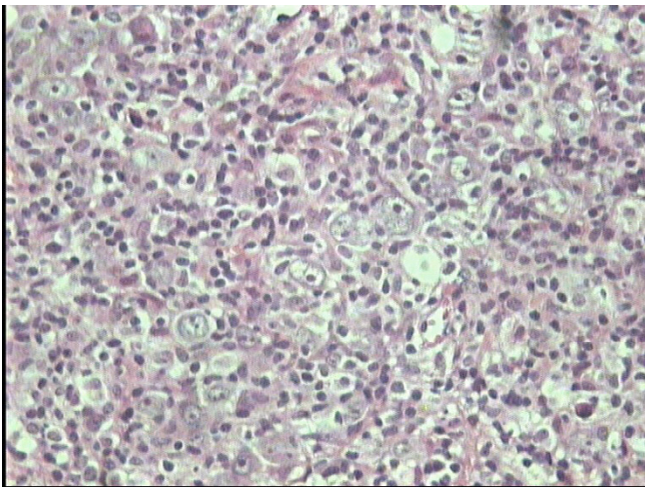


Figure 1
MCHL H/RS cells in an inflammatory background, H&E × 40.

revealed an extraordinarily robust gene expression relationship between PMBL and HL, strongly supporting a pathogenetic relationship between these two lymphoma types [24,25]. On IHC basis, PMBL are PAX5/BSAP+, Bob-1+, Oct-2+, PU.1+, Bcl-2+, CD30+, HLA-DR+, Bcl-6+/-, Mum-1+/- [26], markers already identified on CHL with the exception of PU.1, Bob-1 and Oct-2 [13,19,28,27]. In addition, the MAL protein initially a PMBL marker [29], has been identified on H/RS cells from a case of NSHL in the study of Rosenwald et al [24]. Furthermore, PMBL and HL have rearranged Ig genes but lack surface Ig [10,11,24]. Considering these previous findings together with the results of our research, one can conclude that the expres-

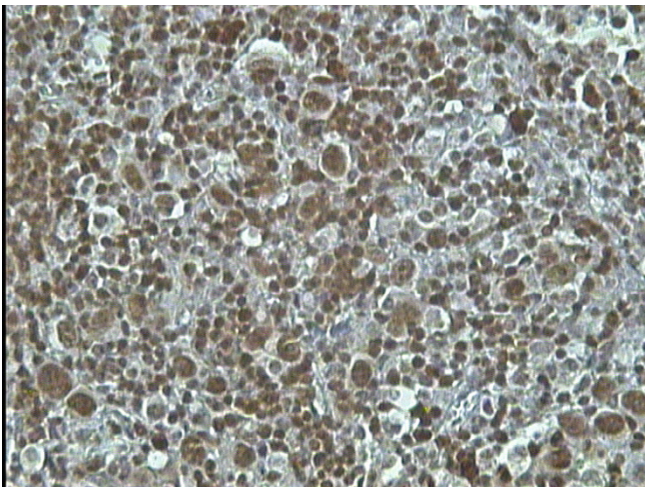


Figure 2
MCHL: strong nuclear Bob-1 staining in all H/RS cells, Bob-1 immunostaining, DAB, Hx, × 40.

Table 2: Bob-1 and EBV expressions in CHL

| CHL Subtype | Bob-1 | EBV viral load | |
|-------------|-------|----------------|-------|
| | | EBER | LMP-1 |
| LRHL | 2/2 | 2/2 | 1/2 |
| MC | 4/4 | 3/4 | 3/4 |
| NS | 9/10 | 6/10 | 6/10 |
| LD | 2/2 | 1/2 | 1/2 |
| Total | 17/18 | 12/18 | 11/18 |
| % | 94% | 67% | 61% |

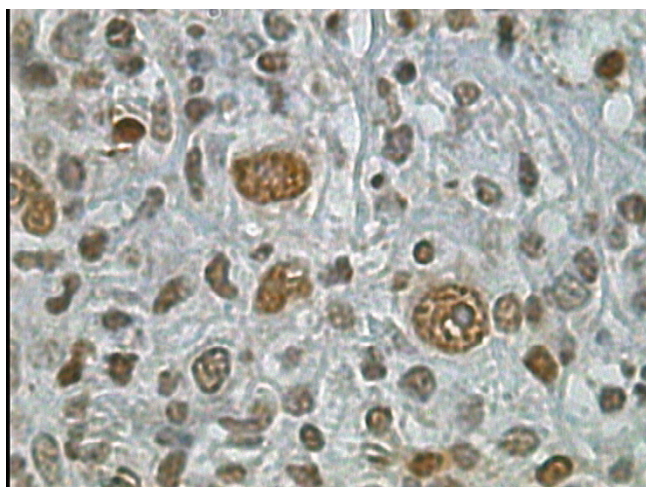


Figure 3
MCHL: higher power magnification of the previous figure, Bob-1 immunostaining, DAB, Hx, $\times 100$.

sion of Bob-1 in the Egyptian population is not an uncommon finding.

What could be the other explanations of Bob-1+H/RS cells in the present study? It has been speculated that EBV contributes to the transformation and maintenance of H/RS cells, by rescuing them from apoptosis. This has been attributed to the oncogenic potential of LMP-1 on B cells through upregulation of anti-apoptosis genes including bcl-2 [30], downregulation of p16INK4a [31] and activation of NF κ B [32]. What is the relation between EBV and Bob-1? NF κ B and Bob-1 are transcription factors required for mouse B cell differentiation, serum IgM production,

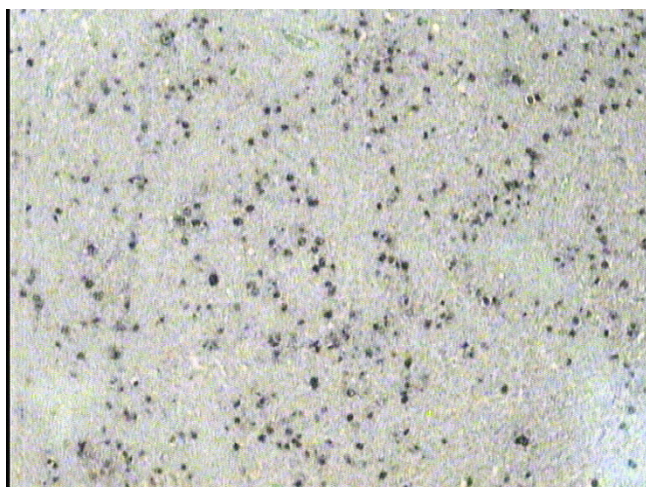


Figure 4
MCHL: strong nuclear EBV staining, EBER-ISH, BNCT, Hx, $\times 20$.

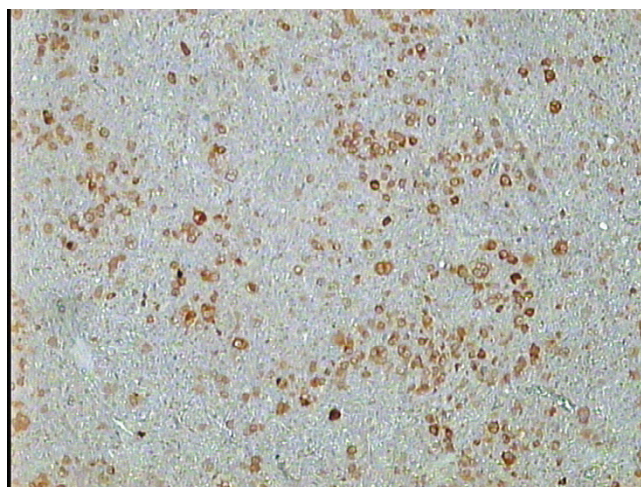


Figure 5
MCHL: LMP-1 immunostaining of the same case, DAB, Hx, $\times 20$.

late B cell maturation and function [33]. It seems that the activation of NF κ B in EBV positive cases is associated with up regulation of Bob-1 since there is a close association between the expression of Bob-1 and EBV in the present study.

In conclusion, this study confirms the nuclear expression of Bob-1 on H/RS cells in CHL, making the utility of Bob-1 to differentiate between LPHL and CHL difficult. These points open questions concerning environmental factors especially early EBV infection in developing countries and to lesser extent genetic ones. Is there truly a difference between Western and Egyptian CHL, or no differences actually exist as most researchers are more concentrated on their own Western-based populations? We feel that our work necessitates cooperative studies between different countries to answer these questions.

Abbreviations

H/RS: Hodgkin/Reed-Sternberg, CHL: classic Hodgkin lymphoma, LPHL: lymphocyte predominance Hodgkin lymphoma, LR: lymphocyte rich, MC: mixed cellularity, NS: nodular sclerosis, LD: lymphocyte depletion, EBV: Epstein-Barr virus, EBER ISH: EBV in situ hybridization, LMP-1: EBV latent membrane protein-1

Competing interests

The author(s) declare that they have no competing interests.

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References

- Jaffe ES, Harris NL, Vardiman JW: **Pathology and Genetics. Tumors of hematopoietic and lymphoid tissues.** WHO Classification of Tumors 2001.
- Galieque Zoutina S, Quief S, Hildebrand MP, Denis C, Lecocq G, Collyn-d'Hooghe M, Bastard C, Yuille M, Dyer MJ, Kerckaert JP: **The B cell transcriptional coactivator BOB1/OBF1 gene fuses to the LAZ3/BCL6 gene by t(3;11)(q27;q23.1) chromosomal translocation in a B cell leukemia line (Karpas 231).** *Leukemia* 1996, **10**:579-587.
- Strubin M, Newell JW, Matthias P: **OBF-1, a novel B cell-specific coactivator that stimulates immunoglobulin promoter activity through association with octamer-binding proteins.** *Cell* 1995, **80**:497-506.
- Schubart DB, Rolink A, Kosco-Vilbois MH, Botteri F, Matthias P: **B-cell-specific coactivator OBF-1/OCA-B/BOB1 required for immune response and germinal centre formation.** *Nature* 1996, **383**:538-542.
- Herbst H, Niedobitek G, Kneba M, Hummel M, Finn T, Anagnostopoulos I, Bergholz M, Krieger G, Stein H: **High incidence of Epstein-Barr virus genomes in Hodgkin's disease.** *Am J Pathol* 1990, **137**:13-18.
- Glaser SL, Lin RJ, Stewart SL, Ambinder RF, Jarrett RF, Brousset P, Pallesen G, Gulley ML, Khan G, O'Grady J, Hummel M, Preciado MV, Knecht H, Chan JK, Claviez A: **Epstein-Barr virus-associated Hodgkin's disease: Epidemiologic characteristics in international data.** *Inter J Cancer* 1997, **70**:375-382.
- Lauritzen AF, Hording U, Nielsen HW: **Epstein-Barr virus, and Hodgkin's disease: a comparative immunological, in situ hybridization, and polymerase chain reaction study.** *APMIS* 1994, **102**:495-500.
- Cossman J, Annunziata CM, Barash S, Staudt L, Dillon P, He WW, Ricciardi-Castagnoli P, Rosen CA, Carter KC: **Reed-Sternberg cell genome expression supports a B cell lineage.** *Blood* 1999, **94**:411-416.
- Foss HD, Reusch R, Demel G, Lenz G, Anagnostopoulos I, Hummel M, Stein H: **Frequent expression of the B cell-specific activator protein in Reed-Sternberg cells of classical Hodgkin's disease provides further evidence for its B cell origin.** *Blood* 1999, **94**:3108-3113.
- Marafioti T, Hummel M, Foss HD, Laumen H, Korbjuhn P, Anagnostopoulos I, Lammert H, Demel G, Theil J, Wirth T, Stein H: **Hodgkin and Reed-Sternberg cells represent an expansion of a single clone originating from a germinal center B cell with functional immunoglobulin gene rearrangements but defective immunoglobulin transcription.** *Blood* 2000, **95**:1443-1450.
- Küppers R, Rajewsky K, Zhao M, Simons G, Laumann R, Fischer R, Hansmann M: **Hodgkin Disease: Hodgkin and Reed-Sternberg cells picked from histological sections show clonal immunoglobulin gene rearrangements and appear to be derived from B cells at various stages of development.** *PNAS* 1994, **91**:10962-10966.
- Kanzler H, Küppers R, Hansmann ML, Rajewsky K: **Hodgkin and Reed-Sternberg cells in Hodgkin's disease represent the outgrowth of a dominant tumor clone derived from (crippled) germinal center B cells.** *J Exp Med* 1996, **184**:1495-1505.
- Stein H, Marafioti T, Foss HD, Laumen H, Hummel M, Anagnostopoulos I, Wirth T, Demel G, Falini B: **Down-regulation of BOB.1/OBF.1 and Oct2 in classical Hodgkin disease but not in lymphocyte predominant Hodgkin disease correlates with immunoglobulin transcription.** *Blood* 2001, **97**:496-501.
- Hertel CB, Zhou XG, Hamilton-Dutoit SJ, Junker S: **Loss of B cell identity correlates with loss of B cell-specific transcription factors in Hodgkin/Reed-Sternberg cells of classical Hodgkin lymphoma.** *Oncogene* 2002, **21**:4908-4920.
- Ushmorov A, Ritz O, Hummel M, Leithauser F, Moller P, Stein H, Wirth T: **Epigenetic silencing of the immunoglobulin heavy-chain gene in classical Hodgkin lymphoma-derived cell lines contributes to the loss of immunoglobulin expression.** *Blood* 2004, **104**:3326-3334.
- Greiner A, Mülle KB, Hess J, Pfeffer K, Müller-Hermelink HK, Wirth T: **Up-Regulation of BOB.1/OBF.1 Expression in Normal Germinal Center B Cells and Germinal Center-Derived Lymphomas.** *Am J Pathol* 2000, **156**:501-507.
- Zwilling S, Dieckmann A, Pfisterer P, Angel P, Wirth T: **Inducible expression and phosphorylation of coactivator BOB.1/OBF.1 in T cells.** *Science* 1997, **277**:221-225.
- Re D, Müschen M, Ahmadi T, Wickenhauser C, Staratschek-Jox A, Holtick U, Diehl V, Wolf J: **Oct-2 and Bob-1 Deficiency in Hodgkin and Reed Sternberg Cells.** *Cancer Res* 2001, **61**:2080-2084.
- Browne P, Petrosyan K, Hernandez A, Chan JA: **The B-Cell transcription factors BSAP, Oct-2, and BOB.1 and the Pan-B-cell markers CD20, CD22, and CD79a are useful in the differential diagnosis of classic Hodgkin Lymphoma.** *Am J Clin Pathol* 2003, **120**:767-777.
- Loddenkemper C, Anagnostopoulos I, Hummel M, Johrens-Leder K, Foss HD, Jundt F, Wirth T, Dorken B, Stein H: **Differential Emu enhancer activity and expression of BOB.1/OBF.1, Oct2, PU.1, and immunoglobulin in reactive B-cell populations, B-cell non-Hodgkin lymphomas, and Hodgkin lymphomas.** *J Pathol* 2004, **202**:60-69.
- García-Cosío M, Santón A, Martín P, Camarasa N, Montalbán C, García JF, Bellas C: **Analysis of transcription factor OCT.1, OCT.2 and BOB.1 expression using tissue arrays in classical Hodgkin's lymphoma.** *Mod Pathol* 2004, **17**:1531-1538.
- Bargou RC, Leng C, Krappmann D, Emmerich F, Mapara MY, Bommert K, Royer HD, Scheidereit C, Dorken B: **High-level nuclear NF-kappa B and Oct-2 is a common feature of cultured Hodgkin/Reed-Sternberg cells.** *Blood* 1996, **87**:4340-4347.
- Joos S, Kupper M, Ohl S, von Bonin F, Mechttersheimer G, Bentz M, Marynen P, Moller P, Pfeundschtuh M, Trumper L, Lichter P: **Genomic imbalances including amplification of the tyrosine kinase gene JAK2 in CD30+ Hodgkin cells.** *Cancer Res* 2000, **60**:549-552.
- Rosenwald A, Wright G, Leroy K, Yu X, Gaulard P, Gascoyne RD, Chan WC, Zhao T, Haioun C, Greiner TC, Weisenburger DD, Lynch JC, Vose J, Armitage JO, Smeland EB, Kvaloy S, Holte H, Delabie J, Campo E, Montserrat E, Lopez-Guillermo A, Ott G, Muller-Hermelink HK, Connors JM, Braziel R, Grogan TM, Fisher RI, Miller TP, LeBlanc M, Chiorazzi M, Zhao H, Yang L, Powell J, Wilson WH, Jaffe ES, Simon R, Klausner RD, Staudt LM: **Molecular diagnosis of primary mediastinal B cell lymphoma identifies a clinically favorable subgroup of diffuse large B cell lymphoma related to Hodgkin lymphoma.** *J Exp Med* 2003, **198**:851-862.
- Savage K, Monti S, effery Kutok JL, Cattoretti G, Neuberg D, de Leval L, Kurtin P, Paola Cin PD, Ladd C, Feuerhake F, Aguiar RCT, Li S, Salles S, Berger F, Jing W, Pinkus GS, Habermann T, Dalla-Favera R, Harris NL, Aster JC, Golub TR, and Margaret A, Shipp MA: **The molecular signature of mediastinal large B-cell lymphoma differs from that of other diffuse large B-cell lymphomas and shares features with classical Hodgkin lymphoma.** *Blood* 2003, **102**:3871-3879.
- Pileri SA, Zinzani PL, Gaidano G, Falini B, Gaulard P, Zucca E, Sabattini E, Ascani S, Rossi M, Cavalli F, International Extranodal Lymphoma Study Group: **Pathobiology of primary mediastinal B-cell lymphoma.** *Leuk Lymphoma* 2003, **44**:S21-S26.
- Carbone A, Gloghini A, Gaidano G, Franceschi S, Capello D, Drexler HG, Falini B, Dalla-Favera R: **Expression status of BCL-6 and syndecan-1 identifies distinct histogenetic subtypes of Hodgkin's disease.** *Blood* 1998, **92**:2220-2228.
- Torlakovic E, Tierens A, Dang HD, Delabie J: **The transcription factor PU.1, necessary for B-cell development, is expressed in lymphocyte predominance, but not classical Hodgkin's disease.** *Am J Pathol* 2001, **159**:1807-1814.
- Copie-Bergman C, Plonquet A, Alonso MA, Boulland ML, Marquet J, Divine M, Moller P, Leroy K, Gaulard P: **MAL expression in lymphoid cells: further evidence for MAL as a distinct molecular marker of primary mediastinal large B-cell lymphomas.** *Mod Pathol* 2002, **15**:1172-1180.
- Rowe M, Peng-Pilon M, Huen DS, Hardy R, Croom-Carter D, Lundgren E, Rickinson AB: **Upregulation of bcl-2 by the Epstein-Barr virus latent membrane protein LMP1: a B-cell-specific response that is delayed relative to NF-kappa B activation and to induction of cell surface markers.** *J Virol* 1994, **68**:5602-5612.
- Yang X, He Z, Xin B, Cao L: **LMP1 of Epstein-Barr virus suppresses cellular senescence associated with the inhibition of p16INK4a expression.** *Oncogene* 2000, **19**:2002-2013.
- Izumi KM, Kieff ED: **The Epstein-Barr virus oncogene product latent membrane protein 1 engages the tumor necrosis factor receptor-associated death domain protein to mediate B lymphocyte growth transformation and activate NF-kB.** *PNAS* 1997, **94**:12592-12597.
- Kim U, Gunther CS, Roeder RG: **Genetic Analyses of NFKB1 and OCA-B Function: Defects in B Cells, Serum IgM Level, and Antibody Responses in Nfkb1-/Oca-b-/ Mice.** *J Immunol* 2000, **165**:6825-6832.