CASE REPORT

Open Access

Primary lung chordoma: a case report



Naoko Shigeta^{1*}, Tetsuya Isaka¹, Kyoko Ono², Mio Tanaka³, Tomoyuki Yokose², Hiroyuki Adachi¹, Wataru Usuba⁴ and Hiroyuki Ito¹

Abstract

Background Chordoma, a rare malignant tumor arising from notochordal tissue, usually occurs along the spinal axis. Only a few published reports of primary lung chordomas exist. Herein, we present a case of primary lung chordoma and discuss important considerations for diagnosing rare chordomas.

Case presentation We report a case of primary lung chordoma in a 39-year-old male with a history of testicular mixed germ-cell tumor of yolk sac and teratoma. Computed tomography revealed slow-growing solid lesions in the left lower lobe. We performed wedge resection for suspected germ-cell tumor lung metastasis. Histologically, large round or oval cells with eosinophilic cytoplasm were surrounded by large cells with granular, lightly eosinophilic cytoplasm. Tumor cells were physaliphorous. Immunohistochemistry was positive for brachyury, S-100 protein, epithelial membrane antigen, vimentin, and cytokeratin AE1/AE3, suggesting pulmonary chordoma. Re-examination of the testicular mixed germ-cell tumor revealed no notochordal elements. Although some areas were positive for brachyury staining, hematoxylin and eosin (HE) staining did not show morphological features typical of chordoma. Complementary fluorescence in situ hybridization (FISH) of the lung tumor confirmed the absence of isochromosome 12p and 12p amplification. Thus, a final diagnosis of primary lung chordoma was established.

Conclusions In patients with a history of testicular mixed germ cell tumors, comparison of histomorphology using HE and Brachyury staining of lung and testicular tumors, and analyzing isochromosome 12p and 12p amplification in lung tumors using FISH is pivotal for the diagnosis of rare lung chordomas.

Keywords Brachyury, Lung chordoma, Testicular mixed germ-cell tumor, Isochromosome 12p, FISH

Background

Chordomas are rare malignant tumors that arise from the remnants of the embryonic notochord. They usually occur along the spinal axis, with most tumors arising in the sacrococcygeal region (50%), spheno-occipital region (30%), or throughout the vertebrae (20%) [1]. Extra-axial chordomas, which arise at a site distant from the notochord, are extremely rare and have been reported in the ulna, tibia, pelvis, deep soft tissue of the knee, gluteus maximus, and posterior chest wall [2]. Herein, we describe a case of primary lung chordoma and discuss important considerations for diagnosing rare chordomas.

Case presentation

An asymptomatic 39-year-old male presented to our department for surgical resection of an 8 mm pulmonary nodule in the left lower lobe of the lung. He had a history of testicular mixed germ cell tumor comprising yolk sac and teratoma components when he was 20 years old. He also had multiple lung, liver, and bone metastases, and elevated anti-human alpha fetoprotein (AFP) (21,500 ng/



© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.gr/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.gr/licenses/by/4.0/.

^{*}Correspondence:

Naoko Shigeta

e123048c@gmail.com

¹ Department of Thoracic Surgery, Kanagawa Cancer Center, 2-3-2 Nakao, Asahi, Yokohama, Kanagawa 241-8515, Japan

² Department of Pathology, Kanagawa Cancer Center, 2-3-2 Nakao, Asahi, Yokohama, Kanagawa 241-8515, Japan

³ Department of Pathology, Kanagawa Children's Medical Center, 1-138-4 Mutsukawa, Minami, Yokohama, Kanagawa 232-0066, Japan

⁴ Department of Urology, St. Marianna University Yokohama Seibu

Hospital, 1197-1 Yasashicyo, Asahi, Yokohama, Kanagawa 241-0811, Japan

mL) levels. The patient underwent inguinal orchiectomy and postoperative chemotherapy with three cycles of cisplatin, etoposide, and bleomycin (BEP). The patient achieved a complete response: all metastatic lesions disappeared on computed tomography (CT) imaging, AFP levels normalized after treatment, and no further treatment for the germ-cell tumor was required. Six years later, a 3 mm pulmonary nodule was detected in the left lower lobe (Fig. 1a). The nodule gradually increased in size, reaching 8 mm after 13 years (Fig. 1a–d). We performed a wedge resection using video-assisted thoracoscopic surgery (VATS) for suspected lung metastasis of a germ-cell tumor. Intraoperative findings revealed a white, jelly-like tumor (Fig. 1e).

Macroscopically, the tumor was a well-defined milkywhite, jelly-like nodule of $8 \times 5 \times 8$ mm, located just below the pleura (Fig. 2).

Hematoxylin and eosin (HE) staining of a paraffin section revealed a tumor encapsulated with fibrous tissue that was clearly separated from the alveoli. Large round or oval cells with eosinophilic cytoplasm were surrounded by large cells with granular, lightly eosinophilic cytoplasm (Fig. 3a-c). The tumor exhibited pleural invasion (d,e) and lymphatic invasion (f,g). The tumor cells exhibited prominent cytoplasmic vacuoles resembling physaliphorous cells, and mildly hyperchromatic and pleomorphic nuclei (Fig. 3b, c). Immunohistochemistry was positive for brachyury (Fig. 3h), S-100 protein, epithelial membrane antigen, vimentin, and cytokeratin AE1/AE3. The Ki-67labeling index was 10%. These findings are consistent with chordoma. Alcian Blue-Periodic Acid Schiff (PAS) staining was used to differentiate chordoma from benign notochordal cell tumor (BNCT), and revealed an extracellular myxoid matrix not seen in BNCT (Fig. 3i).

In addition, immunohistochemical analysis of AFP, Sallike protein 4 (SALL4), PLAP, c-kit, and D2-40, performed to differentiate chordoma from lung metastases of germcell tumors, were all negative. Microscopic reevaluation of HE-stained paraffin sections of the testicular mixed germ-cell tumor measuring $10.5 \times 7.5 \times 8$ cm, resected 19 years prior to the lung resection, showed no evidence of chordoma (Fig. 4a–c). Although some areas contained epithelioid cells positive for brachyury, these cells did not morphologically resemble chordoma (Fig. 4d, e).

Furthermore, some testicular tumor cells with a broad eosinophilic cytoplasm may have been chordoma or notochordal cells owing to their HE staining features. however, all such cells were negative for



Fig. 1 The pulmonary nodule, located in the left lower lobe on computer tomography, is a round, solid nodule with a well-defined border. **a** 13 years, **b** 8 years, **c** 3 years, and **d** 2 weeks before surgery. **e** Intraoperative findings of the pulmonary nodule



Fig. 2 The tumor is 8×5×8 mm, well demarcated, white, and jelly-like



Fig. 3 Microscopic examination revealing (**a**) solid sheets of neoplastic cells (×40). The tumor is composed of (**b**) large round or oval cells with eosinophilic cytoplasm surrounded by (**c**) large cells with granular lightly eosinophilic cytoplasm (×400). **d** The tumor exhibits pleural invasion and (**e**) Elastica van Gieson (EVG) staining confirms pleural disruption (×100). **f** The tumor exhibits lymphatic invasion, and (**g**) Podoplanin (D2-40) staining confirms the presence of tumor cells in the lymphatic vessels (×100). **h** Immunohistochemical staining shows the tumor cells, positive for brachyury (×400). **i** Alcian blue-periodic acid-Schiff staining shows a myxoid intercellular matrix, positive for Alcian blue, surrounding the tumor cells (×400)

brachyury. In addition, fluorescence in situ hybridization (FISH) of the lung tumor revealed the absence of isochromosome 12p and 12p amplification, suggesting lung chordoma (Fig. 5). The pulmonary nodule was therefore diagnosed as a primary lung chordoma. The resected specimen had negative margins and did not require additional treatment. The postoperative course was uneventful, with no recurrence at the 20-month postoperative follow-up.



Fig. 4 Microscopic findings of a testicular mixed germ-cell tumor of the yolk sac and teratoma (× 100). **a** In the yolk sac component of the tumor, small cells proliferate in cystic, luminal, and microcystic forms and Schiller-Duval body, which has a central vessel surrounded by tumor cells, is seen. **b** Both the yolk sac and teratoma components are seen. The squamous and glandular epithelium of the teratoma component are seen. **c** In another teratoma component, cartilage, adipocytes, and epithelium, such as gastrointestinal epithelium, are seen. **d** An area positive for brachyury. **e** HE staining of the same area as (**d**). These epithelioid cells are findings derived from teratoma, not chordoma. None of the areas positive for brachyury show notochordal cells with hematoxylin and eosin staining (×400)



Fig. 5 FISH analysis of the lung chordoma. Green signals represent the centromere of chromosome 12, while red signals correspond to the short arm of chromosome 12. The ratio of red to green signals was 1:1. FISH analysis showed no appearance of isochromosome 12p

Discussion and conclusions

Chordomas are locally aggressive, slow-growing tumors that account for 1-4% of all primary malignant bone tumors and are thought to arise from notochordal cells [3, 4]. Chordomas occur mostly along the spinal axis in patients with a median age of 57 years, and are more

common in males [5]. Extra-axial chordomas are rare and, to date, only a few cases of lung chordomas have been reported (Table 1). Extra-axial chordomas, including lung chordomas, are presumed to originate from a notochordal remnant with aberrant migration from the midline or from multipotent cells in the lung parenchyma Table 1 Departed cases of primary lung chardenay

Table T Reported cases of primary rang chordomas				
Case (ref.)	Age/sex	Symptoms	Location in the	Number

Size Treatment Prognosis lung of nodules 1 [6] 79/F Persistent cough Right lower lobe 1 20 mm Wedge resection No recurrence for 24 and minimal rightmonths sided chest pain 2 [8] 79/M Intermittent fever Right lower lobe 1 73 mm None (Diagnosed None stated with CT-quided and dyspnea biopsy) 3 [9] 40/M None Right lower lobe, left 8 4-23 mm Wedge resection of 2 No recurrence apex and left lingular nodules and no enlargement segment of residual lesion for 38 months 4 [10] Right apex and left 3 No recurrence for 8 61/F None Right, 18 mm; tumor Segmentectomy lower lobe size of left lobe for 3 all nodules months is unknown 5 [11] 59/M Dull, exertional, **Right** apex 36 mm Wedge resection None stated and proton therapy episodic, substernal and right-sided chest for positive margin pain Our case 39/M None Left lower lobe 1 8 mm Wedge resection No recurrence for 4 months

[2, 6]. Kikuchi et. al. suggested that this hypothesis may also apply to the origin of extra-axial benign notochordal cell tumors (BNCTs). BNCTs could potentially serve as a precursor lesion not only for conventional axial chordomas but also for extra-axial chordoma [7].

Macroscopically, lung chordomas are usually welldemarcated, solid to cystic, with a yellow, transparent, gelatinous appearance [9]. Microscopically, HE staining of tumor sections show that intraosseous chordoma cells float in sheets, cords, or alone in an abundant myxoid stroma surrounded by a fibrous band. These cells, known as physaliphorous cells, display a vacuolated cytoplasm, and nuclei were mild-to-moderate atypia [12]. The Ki-67 labeling index in the present case (10%) is consistent with those in previous studies, ranging from 1 to 50% (median, 5%) [13]. Furthermore, chordomas are positive for brachyury, cytokeratin AE1/AE3, epithelial membrane antigen, vimentin, and S-100 protein [12, 14]. Brachyury regulates the development of notochordal cell differentiation, and positive immunohistochemistry of brachyury can differentiate chordomas from chondrosarcomas and chordoid gliomas [7]. However, benign notochordal cell tumors (BNCT) are also positive for brachyury. Several studies have attempted to distinguish chordomas from BNCT. According to these prior studies, unlike BNCT, tumor cells in chordomas have a myxoid matrix between them [7, 15]. In our case, Alcian blue-PAS staining revealed an abundant myxoid intercellular matrix. Second, according to the studies, a chordoma is encompassed by a thin fibrous membrane with a very smooth border, while a BNCT does not show any fibrous capsule formation [15]. In our case, the tumor was encapsulated by a fibrous tissue that was separated from the alveoli. Third, a previous study suggested that the nuclei of chordoma are much more atypical than those of BNCT [15]. In our case, the nuclei were mildly hyperchromatic and pleomorphic. Thus, we considered the tumor a chordoma rather than BNCT. In addition, the malignant findings of pleural and lymphatic invasion supported the diagnosis of chordoma.

Previous reports of lung chordomas have described microscopic findings similar to those of chordomas arising from notochordal tissue. The tumor cells of lung chordomas are surrounded by a fibrous capsule that separates them from the surrounding alveoli [16]. Furthermore, immunohistochemistry of lung chordomas has been reported to be positive for the same antibodies as chordomas arising from notochordal tissue [6, 8-10]. The microscopic and macroscopic findings of lung chordomas presented in this study were consistent with those previously reported. Previously published reports on lung chordomas are summarized in Table 1.

The median age was 61 (49-79) years, and the tumors ranged in size from 4 to 73 mm and were often round and solid (Table 1). Our case represents the youngest reported case, both in terms of the age at discovery and at surgery. Apart from our case study, only Ohya et al. reported long-term CT follow-up of primary lung chordoma, involving a 40-year-old patient exhibiting multiple tumors that gradually increased in size over a 14-year period [3]. In our study, the solitary tumor

grew slowly and increased in diameter by 5 mm over a period of 13 years. A diagnosis of primary chordoma should therefore be considered when a solitary tumor is slow-growing and has a round, solid appearance on CT in young patients.

Careful differentiation should be made between primary lung chordomas and metastatic lung tumors of germ-cell tumors because some testicular germ-cell tumors are known to express brachyury protein [16]. Studies have reported that brachyury nuclear staining was observed in 23 out of 96 (24%) testicular germ cell tumors and it was most frequently observed in mixed tumors along with teratomas (33.3%), followed by mixed tumors (25.9%) [17]. In addition, a teratoma containing a chordoma component has been reported in a case of ovarian teratoma [18]. This case required special attention to determine whether the brachyury-positive cells found in the germ cell tumor were cells of a germ cell origin or chordoma component. This case study successfully differentiated primary lung chordoma from a metastatic lung germ-cell tumor using four methodologies. Firstly, we confirmed the absence of 12p abnormalities by FISH analysis in the resected lung chordoma. Isochromosome 12p and 12p amplification are fundamental abnormalities associated with germ cell tumors and holds significant diagnostic value in their identification [19]. Germ cell tumors can be categorized into two types: postpubertal type with 12p abnormalities, arising from germ cell neoplasia in situ (GCNIS), and prepubertal type without 12p abnormalities, developing without GCNIS [20]. Prepubertal-type teratomas typically occur before the age of 6 years, and prepubertal-type yolk sac tumors are found in 2 to 3 cases per million children aged 0-5 years [21]. Given that the germ cell tumor occurred at 20 years of age, it was considered post-pubertal type with 12p abnormalities. In cases with 12p amplification, FISH analyses show more red signals corresponding the short arm of chromosome 12 in relation to green signals corresponding the centromere of chromosome 12. In cases with isochromosome 12p, FISH analyses show one green signal and two red signals in close proximity because the short arms of the chromosomes are close to each other. In this case of lung chordoma, the ratio of green signals to red signals were one to one in FISH analysis, and neither 12p amplification nor isochromosome 12p was not observed, making metastasis of the germ cell tumor unlikely. Second, careful reexamination of the testicular mixed germ-cell tumor revealed no chordoma component throughout the germ-cell tumor. Third, HE staining showed no morphological structures characteristic of chordoma in the areas where Brachyury staining was positive. Fourth, immunohistochemical analysis of the pulmonary nodule for germ-cell markers, including AFP, SALL4, PLAP, c-kit, and D2-40 was negative, making testicular germ-cell tumor metastasis unlikely.

The primary treatment of chordoma is surgery. No effective chemotherapy has been identified. Radiation therapy may improve local control and delay disease progression in some patients in whom complete resection is not possible [22]. McMaster et al. reported that the median survival of 361 patients with a primary chordoma was 6.29 years, with overall 5- and 10-year relative survival rates of 67.6% and 39.9%, respectively [23]. According to published reports of primary lung chordoma, four out of five cases of primary lung chordoma underwent surgery, whereas one patient refused surgery and was therefore followed up without treatment (Table 1). In our case, the tumor was completely resected using wedge resection under VATS, and there was no recurrence within 20 months postoperatively.

In conclusion, we performed complete resection of a rare primary lung chordoma that appeared as a slowgrowing, solid lung tumor on CT in a patient with a history of testicular germ-cell tumor. FISH analysis of isochromosome 12p and 12p amplification of lung tumors and comparison of pathomorphology of lung and testicular tumors by HE staining and Brachyury staining are crucial in differentiating lung chordoma from metastatic lung cancer of testicular germ cell origin.

Abbreviations

 FISH
 Fluorescence in situ hybridization

 CT
 Computed tomography

 BNCT
 Benign notochordal cell tumor

- BNCT Benign notochordal cell tumor AFP Anti-human alpha fetoprotein
- ALL A Callilla anatain A
- SALL4 Sal-like protein 4

Acknowledgements

We would like to thank Editage (www.editage.com) for English language editing.

Authors' contributions

TI, WU, and NS collected clinical information, diagnostic information, therapeutic information, and images of the patients. NS drafted the manuscript. TI wrote the manuscript. KO, TY, and NS collected the pathological images of the patients. MT performed FISH analysis. TI, KO, TY, HA and HI revised the manuscript. All authors contributed to the manuscript and approved the submitted version.

Funding

None declared.

Availability of data and materials

Not applicable. Our manuscript does not contain any numerical data.

Declarations

Ethics approval and consent to participate

Our institution does not require ethics committee approval for Case Reports. The investigation was conducted in accordance with the Declaration of Helsinki of 1975.

Consent for publication

This case report has consent from patient for publication.

Competing interests

The authors declare no competing interests.

Received: 3 April 2023 Accepted: 26 June 2024 Published online: 03 July 2024

References

- Stacchiotti S, Sommer J, Chordoma Global Consensus G. Building a global consensus approach to chordoma: a position paper from the medical and patient community. Lancet Oncol. 2015;16:e71–83.
- DiFrancesco LM, Davanzo Castillo CA, Temple WJ. Extra-axial chordoma. Arch Pathol Lab Med. 2006;130:1871–4.
- Gulluoglu S, Turksoy O, Kuskucu A, Ture U, Bayrak OF. The molecular aspects of chordoma. Neurosurg Rev. 2016;39:185–96; discussion 96.
- Mirra JM, Nelson SD, Della Rocca C, Mertens F. Chordoma. In: Fletcher CDM, Unni KK, Mertens F, editors. Pathology and genetics of tumors of soft tissue and bone World Health Organization classification of tumors. Lyon: IARC Press; 2002. p. 316–7.
- Zhou J, Sun J, Bai XH, Huang X, Zou Y, Tan X, et al. Prognostic factors in patients with spinal chordoma: an integrative analysis of 682 patients. Neurosurgery. 2017;81:883.
- Strano S, Ouafi L, Baud M, Alifano M. Primary chordoma of the lung. Ann Thorac Surg. 2010;89:302–3.
- Kikuchi Y, Yamaguchi T, Kishi H, Azuhata K, Kimizuka G, Hiroshima K, et al. Pulmonary tumor with notochordal differentiation: report of 2 cases suggestive of benign notochordal cell tumor of extraosseous origin. Am J Surg Pathol. 2011;35:1158–64.
- Park SY, Kim SR, Choe YH, Lee KY, Park SJ, Lee HB, et al. Extra-axial chordoma presenting as a lung mass. Respiration. 2009;77:219–23.
- 9. Ohya M, Yoshida K, Shimojo H, Shiina T. Multiple primary chordomas of the lung. Respir Med Case Rep. 2018;25:142–4.
- Kaufman D, Farias V, Lipton J, Brichkov I. Lung chordoma: a discussion. Ann Thorac Surg. 2022;114:e33–4.
- Ball S, Dash A, Igid HP, Thein KZ, Sharma U, Tijani L. Primary extra-axial chordoma masquerading as lung cancer: case report and review of the literature. Clin Lung Cancer. 2020;21:e560–3.
- Walcott BP, Nahed BV, Mohyeldin A, Coumans JV, Kahle KT, Ferreira MJ. Chordoma: current concepts, management, and future directions. Lancet Oncol. 2012;13:e69–76.
- von Witzleben A, Goerttler LT, Lennerz J, Weissinger S, Kornmann M, Mayer-Steinacker R, et al. In chordoma, metastasis, recurrences, Ki-67 index, and a matrix-poor phenotype are associated with patients' shorter overall survival. Eur Spine J. 2016;25:4016–24.
- 14. Abenoza P, Sibley RK. Chordoma: an immunohistologic study. Hum Pathol. 1986;17:744–7.
- Yamaguchi T, Imada H, Iida S, Szuhai K. Notochordal tumors: an update on molecular pathology with therapeutic implications. Surg Pathol Clin. 2017;10:637–56.
- Tirabosco R, Mangham DC, Rosenberg AE, Vujovic S, Bousdras K, Pizzolitto S, et al. Brachyury expression in extra-axial skeletal and soft tissue chordomas: a marker that distinguishes chordoma from mixed tumor/myoepithelioma/parachordoma in soft tissue. Am J Surg Pathol. 2008;32:572–80.
- Pinto F, Carcano FM, da Silva ECA, Vidal DO, Scapulatempo-Neto C, Lopes LF, et al. Brachyury oncogene is a prognostic factor in high-risk testicular germ cell tumors. Andrology. 2018;6:597–604.
- Las Heras F, Pritzker KP, Colgan TJ. Chordoma arising in a mature cystic teratoma of the ovary: a case report. Pathol Res Pract. 2007;203:467–71.
- Freitag CE, Sukov WR, Bryce AH, Berg JV, Vanderbilt CM, Shen W, et al. Assessment of isochromosome 12p and 12p abnormalities in germ cell tumors using fluorescence in situ hybridization, single-nucleotide polymorphism arrays, and next-generation sequencing/mate-pair sequencing. Hum Pathol. 2021;112:20–34.
- 20. Iczkowski KA. Germ cell neoplasms of the testis: update for 2022. Semin Diagn Pathol. 2023;40:2–21.
- Ulbright TM, Amin MB, Balzed B, Berney DM, Epstein JI, Guo C, et. al. Germ cell tumors. In: World Health Organization classification of the urinary system and male genital organs. 4th ed. Lyon: World Health Organization; 2016. p. 189–226.

- 22. Bergh P, Kindblom LG, Gunterberg B, Remotti F, Ryd W, Meis-Kindblom JM. Prognostic factors in chordoma of the sacrum and mobile spine: a study of 39 patients. Cancer. 2000;88:2122–34.
- McMaster ML, Goldstein AM, Bromley CM, Ishibe N, Parry DM. Chordoma: incidence and survival patterns in the United States, 1973–1995. Cancer Causes Control. 2001;12:1–11.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.